

Leptospira spp. and *Dirofilaria immitis* in dogs in Tonga: a pilot study

Dogs are common in Tonga and exist in a close relationship with humans, both as free-ranging pets and guard dogs for domestic properties. Little is known about pathogens present in these animals, and this pilot study aimed to develop a methodology to identify the presence of zoonotic pathogens *Leptospira* spp. and *Dirofilaria immitis* (the causative agent of heartworm), in a sample of 82 dogs voluntarily presented to a spay/neuter clinic, using blood samples and point-of-care tests. No positive tests were returned for *Leptospira* spp. or *D. immitis*, despite the presence of *Leptospira* spp. having previously been identified in Tonga.

Kristina Naden BVN RVN NZCATT, Lecturer, School of Veterinary Nursing, Otago Polytechnic, Dunedin, New Zealand; **Kate Harder** BSc(Marine Ecology) PG CertSciTech(Conservation Biology) RVN NZCATT, Lecturer, School of Environmental and Animal Sciences, Unitec Institute of Technology, Auckland, New Zealand. kristina.naden@op.ac.nz

Key words: Tonga | *Leptospira* | *Dirofilaria immitis* | heartworm | canine

The Kingdom of Tonga's main island of Tongatapu has a human population of 74 611 (Tonga Department of Statistics, 2016), and an estimated canine population of 17 122 dogs, giving a dog:human ratio of 1:4 (G Aguilar, unpublished data, 2018), similar to other developing countries such as Nepal, Indonesia and the Philippines (Jackman and Rowan, 2007; Traub et al, 2015). The close physical relationship that dogs have with humans in Tonga, as both pets and guard dogs, inherently increases the risk of disease transmission, since more than 60% of known infectious diseases are zoonotic (Bidaisee and Macpherson, 2014).

This pilot study considers two specific zoonotic pathogens – *Leptospira* spp. and *Dirofilaria immitis* – in dogs in the Kingdom of Tonga. Leptospirosis has been identified in 13 Pacific nations, including Tonga (Guernier et al, 2018). However, under-reporting of this disease is common, and it is known to mimic other diseases (Izurieta et al, 2008; Victoriano et al, 2009). Cattle, swine, dogs and rats are common reservoirs for *Leptospira* spp., and shed leptospires in urine. In countries with tropical climates, such as the Philippines and Thailand, 68–92% of rats were found to carry antibodies for leptospires (Tangkanakul et al, 2005; Villanueva et al, 2010). Dogs infected with *Leptospira* spp. typically present with fever, jaundice, vomiting, diarrhoea, intravascular disseminated coagulation, renal failure, haemorrhages and death. Infection may occur via direct or indirect exposure,

where rivers, soil and water reservoirs may be contaminated by urine of carrier animals (Victoriano et al, 2009; Guernier et al, 2018). Spikes in infection rates in humans are often noted following cyclones with secondary flooding, and projected changes in the world's climate are expected to increase the frequency of these severe weather events (Gubler et al, 2001; Lau et al, 2016). Clinical signs in humans range from a mild flu-like illness, to severe complications including acute renal failure and pulmonary haemorrhagic syndrome. The latter is associated with high fatality rates (Guernier et al, 2018).

Human infections of *D. immitis* are unusual, while transmission of this nematode via the mosquito as a vector have been reported (Mendoza-Roldan et al, 2021). It is considered an emerging zoonotic disease that may be transmitted between humans and carnivores in tropical and subtropical regions (Vieira et al, 2014; Anvari et al, 2020). Clinical signs of heartworm caused by *D. immitis* in dogs include a cough, dyspnoea, congestive heart failure, physical activity intolerance, intravascular haemolysis, haemoptysis, ascites, pulmonary thromboembolism, loss of appetite and weight loss (Anvari et al, 2020). *D. immitis* may be transmitted to humans in its third larval stage, and transmission may cause benign pulmonary nodules in humans, that can be misdiagnosed as pulmonary carcinomas (Simón et al, 2009; Montoya-Alonso et al, 2011; Vieira et al, 2014).

The zoonotic nature of both pathogens in this pilot study highlight the importance of a One Health approach to disease control, where links between animal, human and environmental health are considered simultaneously.

Leptospira spp. and *D. immitis* have a higher incidence in tropical countries (Berlioz-Arthaud, 2003; Simón et al, 2009; Montoya-Alonso et al, 2011; Chadsuthi et al, 2017), and dogs living in conditions similar to those found in Tonga (outdoor lifestyle, exposure to raw sewage, scavenging for food, consumption of raw meat, and the prevalence of vectors such as mosquitos), are at a higher risk of exposure to both *Leptospira* spp. and *D. immitis* (Meeyam et al, 2006; Carslake et al, 2017). An estimated 10.9% of dogs are infected with *D. immitis* globally (Anvari et al, 2020).

The lack of Tonga-specific data on these two pathogens is partially because of the limited access to veterinary services, with many Pacific nations, including Tonga, having no permanent veterinary clinic or qualified veterinary personnel. The Kingdom of Tonga relies on volunteer veterinary surgeons and veterinary nurses to provide treatment for animals in the form of temporary clinics. These typically focus on desexing companion animals, with one such clinic used to construct a dataset on the *Leptospira* spp. and *D. immitis* prevalence in dogs. While this report is the only current systematic analysis of the presence of these two pathogens in Tonga, it is limited because of sample bias. However, it provides a method to be replicated in future studies of dogs, both in Tonga and other Pacific nations.

Materials and methods

Sample collection

Inclusion criteria included a minimum age of 6 months for dogs and admission to the clinic for desexing. To be eligible for desexing, dogs were required to be clinically healthy. The minimum age was selected because *D. immitis* is only detectable in infected dogs aged more than 6 months (Santoro et al, 2019). If age was unknown, the dog was aged dentally and any dogs with deciduous dentition were excluded. Of the dogs admitted for desexing, 82 were eligible to be tested for *Leptospira* spp. antibodies and *D. immitis* antigens, based on the above criteria.

A blood sample was collected via intravenous catheter during anaesthesia for each dog's desexing surgery. Blood samples were placed in labelled, additive-free Vacutainer

tubes, and stored in a standard refrigerator to allow clotting and separation of blood samples. Gender, age (where known) and weight were recorded. Ethics approval for this study was obtained by the AgResearch Animal Ethics Committee, Ruakura, New Zealand, application number 14611. All experiments were conducted in accordance with AgResearch Animal Ethics Committee guidelines.

Testing

To test for the presence of *Leptospira* spp. antibodies and *D. immitis* antigens in situ, canine *Leptospira* antibody SNAP tests and canine heartworm SNAP tests (IDEXX Laboratories, USA) were used in accordance with manufacturer's instructions. To obtain serum, 3 ml of serous blood from each dog's blood sample was extracted via pipette, placed in an Eppendorf tube, and spun at 10 000 RPM in a centrifuge for 3 minutes to separate the serum from red blood cells. Each test required three drops of serum and four drops of conjugate, which were placed in the sample tube provided with each SNAP test. The sample tube was capped and inverted a number of times to thoroughly mix the contents. The entire contents of the sample tube were dropped via pipette into the sample well of *D. immitis* and *Leptospira* spp. SNAP tests and labelled with the dog's identification number. Once the sample had flowed to the activation window, the activator was pressed firmly. Each test was read after the required time, in accordance with package insert instructions. Results were recorded for each dog against their identification number.

Results

A total of 82 dogs, all from Tonga's main island of Tongatapu, met criteria to be tested for both *Leptospira* spp. and *D. immitis*. All dogs tested were of mixed-breed, primarily from the Tokomololo village area, and 48% of dogs tested were juvenile (more than 6 months old, but less than 12 months). Of these, 39 were male and 43 were female (Table 1). None of the dogs tested had been vaccinated for *D. immitis* or *Leptospira* spp.

All 82 dogs in this pilot study tested negative for both *Leptospira* spp. and *D. immitis*.

Discussion

The lack of *D. immitis* and *Leptospira* spp. was unexpected because of prevalence of *D. immitis* has been reported at 22–86% in Asia Pacific countries (Carslake et al, 2017), and

Table 1. Gender and age distribution of dogs tested for prevalence of *Leptospira* spp. and *D. immitis*

Gender	6–12 months old	12–24 months old	24+ months old	Unknown age	Total
Male	15	7	4	13	39
Female	25	6	7	5	43
Total	40	13	11	18	82

Leptospira spp. has previously been identified in swine and cattle in Tonga (Saville, 1996), with serogroups Australis and Pomona found in both species, Hebdomadis, Sejroe and Tarassovi found only in cattle, and Icterohaemorrhagiae found only in swine. The serovars Canicola, Icterohaemorrhagiae, Grippotyphosa, and Pomona are commonly found in dogs (Izurieta et al, 2008). No previous quantitative data for *D. immitis* in Tonga has been identified, but nearly 47% of dogs studied in the neighbouring Pacific nation of Samoa were found to carry *D. immitis* antigens (Carslake et al, 2017).

This pilot study was limited to one species (dogs). Future studies into the presence of *Leptospira* spp. in particular may be expanded to include additional species, such as rats, swine and cattle, as well as the testing of water samples, to further investigate the presence of leptospires. Polymerase chain reaction testing has been successfully used to detect leptospires in water, to test for the presence of the lipL32 protein (Muñoz-Zanzi et al, 2014; Wynwood et al, 2014).

This study was also limited by sample bias and the testing method available. Based on these limitations, and because of the limited data available on the health status of animals in Tonga, further development and expansion of this pilot study is warranted.

Sample population

For both tests, dogs were not randomly selected to be tested at the clinic, thus introducing bias in the sample population. Further studies should include other regions of Tongatapu, as well as other islands of Tonga where possible, to provide access to a geographically wider sample population. Future studies should include dogs that may not ordinarily be presented at a veterinary clinic, including those who are not clinically healthy.

The health of dogs in Tonga is challenged by poor nutrition and limited access to preventative health care (K Harder et al, unpublished data, 2023), meaning that a dog may not survive when challenged by a pathogen such as *Leptospira* spp. or *D. immitis*. Infection and subsequent development of leptospirosis can carry high mortality rates of up to 43.4% (Major et al, 2014), with younger dogs being more susceptible to peracute or acute death (Rissi and Brown, 2014). The sample for this study was selected from dogs presenting for desexing at a temporary spay/neuter clinic set up for 1 week. However, to be eligible for desexing, all dogs underwent a clinical exam to confirm they were clinically healthy. Many dogs in Tonga are not desexed, resulting from the lack of access to veterinary treatment and, anecdotally, negative owner perceptions around the need for desexing. This puts entire male dogs, in particular, at higher risk for infection with *Leptospira* spp. because of their greater roaming and contact with urine (Major et al, 2014).

Testing method

Three methods are commonly used to identify the pres-

'The health of dogs in Tonga is challenged by poor nutrition and limited access to preventative health care'

ence of *Leptospira* spp.: enzyme-linked immunosorbent assay, microscopic agglutination test and polymerase chain reaction test. The latter two require specialised testing equipment and personnel, not available in Tonga. Therefore, microscopic agglutination test and polymerase chain reaction test would require transporting biological samples to a location with suitable facilities and technicians, imposing environmental and border security risks. In addition, microscopic agglutination test testing has been reported as subjective for leptospirosis (Winzelberg et al, 2015), with discordant results shown from various testing laboratories (Miller et al, 2011). The chosen method of enzyme-linked immunosorbent assay using SNAP tests (IDEXX), is commonly used in veterinary clinics as a 'point-of-care' test. Curtis et al (2015) identified a 79.2% overall agreement of SNAP tests with microscopic agglutination testing, with variations based on titre levels. However, alternative enzyme-linked immunosorbent assay tests, in the form of the WITNESS Lepto Rapid Test (Zoetis, USA), have shown improved performance when compared with SNAP tests and microscopic agglutination test (Lizer et al, 2017).

The most common serovars of *L. interrogans* thought to infect dogs, before the introduction of vaccination in the 1980s, were Icterohaemorrhagiae and Canicola (Sykes et al, 2011), and the *L. interrogans* serovar Pomona strain has been previously identified in cattle and swine in Tonga (Saville, 1996). The serogroups Icterohaemorrhagiae and Australis, commonly found in rats, were identified in the western Pacific islands (Victoriano et al, 2009). Serovars in the SNAP tests used are limited to Grippotyphosa, Canicola, Pomona and Icterohaemorrhagiae. However, additional serovars not included in tests used in this pilot study, may be present in Tonga.

While no positive tests were returned in this pilot study, *Leptospira* spp. has previously been detected in livestock in Tonga, and the practice of keeping dogs as pets is becoming more common in developing countries, possibly as a result of increasing urbanisation and wealth (Traub et al, 2015). This increase in both population and interaction between dogs and humans brings possible additional opportunities for disease transmission, with free-roaming dogs more likely to transmit disease (Farnworth et al, 2012).

Conclusions

While this pilot study found no evidence of *Leptospira* spp. or *D. immitis* in the sample of dogs, further studies are warranted, in particular for *Leptospira* spp. because of previous

KEY POINTS

- *Dirofilaria immitis* and *Leptospira* spp. have been identified in Tonga, but no previous studies have been carried out on their presence in dogs.
- Both infectious diseases are zoonotic, which poses a public health risk.
- This pilot study aimed to develop a methodology to identify the presence of both diseases in dogs in Tonga. Future studies are warranted, and a refined method is suggested.

evidence found in Tonga and the global importance of leptospirosis. Expanding sampling options may assist in developing a clearer idea of the presence of these pathogens in Tonga. Environmental sampling of water for evidence of leptospires may identify non-mammalian reservoirs, and sampling of additional hosts such as cattle, swine and rats may identify mammalian reservoirs. **VN**

Conflicts of interest

The authors declare that there are no conflicts of interest.

Acknowledgements

Funding for the study was provided in part by Unitec Institute of Technology. IDEXX Laboratories provided a discount for leptospirosis and heartworm SNAP tests. The authors would like to thank South Pacific Animal Welfare, the Tongan Ministry of Agriculture and Food, Forests and Fisheries, the Tongan Animal Welfare Society, Laura Harvey, Dr Kristie Cameron, Lauren Prior and Dr Robyn Gear, from the School of Environmental and Animal Sciences at Unitec Institute of Technology, for support provided.

References

- Anvari D, Narouei E, Daryani, A et al. The global status of *Dirofilaria immitis* in dogs: a systematic review and meta-analysis based on published articles. *Res Vet Sci*. 2020;131:104–116. <https://doi.org/10.1016/j.rvsc.2020.04.002>
- Berlitz-Arthaud A. Survey on Leptospirosis in the Pacific 2003. <https://www.pphsn.net/wp-content/uploads/2017/12/Survey-lepto-Pacific.pdf> (accessed 3 April 2023)
- Bidaise S, Macpherson CN. Zoonoses and one health: a review of the literature. *J Parasitol Res*. 2014;2014:874345. <https://doi.org/10.1155/2014/874345>
- Carslake R, Hill K, Sjolander K, Hii S, Prattley D, Acke E. A cross-sectional survey of health management and prevalence of vector-borne diseases, endoparasites and ectoparasites in Samoan dogs. *Aust Vet J*. 2017;95(12):462–468. <https://doi.org/10.1111/avj.12647>
- Chadsuthi S, Bicout DJ, Wiratsudakul A et al. Investigation on predominant *Leptospira* serovars and its distribution in humans and livestock in Thailand, 2010–2015. *PLoS Negl Trop Dis*. 2017;11(2):2010–2015. <https://doi.org/10.1371/journal.pntd.0005228>
- Curtis KM, Foster PC, Smith PS et al. Performance of a recombinant LipL32 based rapid in-clinic ELISA (SNAP® Lepto) for the detection of antibodies against *Leptospira* in dogs. *Int J Appl Res Vet Med*. 2015;13(3):182–189
- Farnworth MJ, Blaszkak KA, Hiby EF, Waran NK. Incidence of dog bites and public attitudes towards dog care and management in Samoa. *Animal Welfare*. 2012;21(4):477–486. <https://doi.org/10.7120/09627286.21.4.477>
- Gubler DJ, Reiter P, Ebi KL, Yap W, Nasci R, Patz, JA. Climate variability and change in the United States. *Environ Health Perspect*. 2001;109(Suppl 2):223–233. <https://doi.org/10.1289/ehp.109-1240669>
- Guernier V, Goarant C, Benschop J, Lau CL. A systematic review of human and animal leptospirosis in the Pacific Islands reveals pathogen and reservoir diversity. *PLoS Negl Trop Dis*. 2018;12(5):e0006503. <https://doi.org/10.1371/journal.pntd.0006503>
- Izurieta R, Galwankar S, Clem A. Leptospirosis: The 'mysterious' mimic. *J Emerg Trauma Shock*. 2008;1(1):21–33. <https://doi.org/10.4103/0974-2700.40573>
- Jackman J, Rowan AN. Free-roaming dogs in developing countries: the benefits of capture, neuter, and return programs. In: Salem DJ, Rowan AN (eds). *The state of the animals 2007*. Washington, DC: Humane Society Press;2007;55–78
- Lau CL, Watson CH, Lowry JH et al. Human leptospirosis infection in Fiji: an eco-epidemiological approach to identifying risk factors and environmental drivers for transmission. *PLoS Negl Trop Dis*. 2016;10(1):1–25. <https://doi.org/10.1371/journal.pntd.0004405>
- Lizer J, Velineni S, Weber A, Krecic M, Meeus P. Evaluation of 3 serological tests for early detection of *Leptospira*-specific antibodies in experimentally infected dogs. *J Vet Intern Med*. 2017; 32:201–207. <https://doi.org/10.1111/jvim.14865>
- Major A, Schweighauser A, Francey T. Increasing incidence of canine leptospirosis in Switzerland. *Int J Environ Res Public Health*. 2014;11(7):7242–7260. <https://doi.org/10.3390/ijerph110707242>
- Meeyam T, Tablerk P, Petchanok B, Pichpol D, Padungtod P. Seroprevalence and risk factors associated with leptospirosis in dogs. *Southeast Asian J Trop Med Public Health*. 2006;37(1):148–153
- Mendoza-Roldan J A, Gabrielli S, Cascio A et al. Zoonotic *Dirofilaria immitis* and *Dirofilaria repens* infection in humans and an integrative approach to the diagnosis. *Acta Tropica*. 2021; 223:106083. <https://doi.org/10.1016/j.actatropica.2021.106083>
- Miller MD, Annis KM, Lappin MR, Lunn KF. Variability in results of the Microscopic Agglutination Test in Dogs with clinical leptospirosis and dogs vaccinated against leptospirosis. *J Vet Intern Med*. 2011;25:426–432. <https://doi.org/10.1111/j.1939-1676.2011.0704.x>
- Montoya-Alonso JA, Carretón E, Corbera JA et al. Current prevalence of *Dirofilaria immitis* in dogs, cats and humans from the island of Gran Canaria, Spain. *Vet Parasitol*. 2011;176(4):291–294. <https://doi.org/10.1016/j.vetpar.2011.01.011>
- Muñoz-Zanzi C, Mason MR, Encina C, Astroza A, Romero A. *Leptospira* contamination in household and environmental water in rural communities in southern Chile. *Int J Environ Res Public Health*. 2014;11(7):6666–80. <https://doi.org/10.3390/ijerph110706666>
- Rissi DR, Brown CA. Diagnostic features in 10 naturally occurring cases of acute fatal canine leptospirosis. *J Vet Diagn Invest*. 2014;26(6):799–804. <https://doi.org/10.1177/1040638714553293>
- Santoro M, Miletti G, Vangone L, Spadari L, Reccia S, Fusco G. Heartworm disease (*Dirofilaria immitis*) in two roaming dogs from the urban area of Castel Volturno, southern Italy. *Front Vet Sci*. 2019;6:1–4. <https://doi.org/10.3389/fvets.2019.00270>
- Saville P. The animal health status of Tonga. 1996. https://www.spc.int/DigitalLibrary/Doc/LRD/AHP/Animal_Health/18678_1996_Animal_Health_Tonga.pdf (accessed 3 April 2023)
- Simón F, Morchón R, González-Miguel J, Marcos-Atxutegi C, Siles-Lucas M. What is new about animal and human dirofilariosis? *Trends Parasitol*. 2009;25(9):404–409. <https://doi.org/10.1016/j.pt.2009.06.003>
- Sykes JE, Hartmann K, Lunn KF, Moore GE, Stoddard RA, Goldstein RE. 2010 ACVIM small animal consensus statement on Leptospirosis: diagnosis, epidemiology, treatment, and prevention. *J Vet Intern Med*. 2011;25(1):1–13. <https://doi.org/10.1111/j.1939-1676.2010.0654.x>
- Tangkanakul W, Smits HL, Jatanasen S, Ashford DA. Leptospirosis: an emerging health problem in Thailand. *Southeast Asian J Trop Med Public Health*. 2005;36(2):281–288
- Tonga Department of Statistics. Tonga statistics at a glance. In: *Census of Population and Housing*. 2016. <https://tongastats.gov.to/census-2/population-census-2/> (accessed 3 April 2023)
- Traub RJ, Irwin P, Dantas-Torres F et al. Toward the formation of a companion animal parasite council for the tropics (CAPCT). *Parasit Vectors*. 2015; 8:271. <https://doi.org/10.1186/s13071-015-0884-4>
- Victoriano AFB, Smythe HL, Gloriani-Barzaga N et al. Leptospirosis in the Asia Pacific region. *BMC Infect Dis*. 2009;9:147. <https://doi.org/10.1186/1471-2334-9-147>
- Vieira AL, Vieira MJ, Oliveira JM, Simoes AR, Diez-Banos P, Gestal J. Prevalence of canine heartworm (*Dirofilaria immitis*) disease in dogs of central Portugal. *Parasite*. 2014;21:5. <https://doi.org/10.1051/parasite/2014003>
- Villanueva SYAM, Ezoe H, Baterna RA et al. Serologic and molecular studies of *Leptospira* and leptospirosis among rats in the Philippines. *Am J Trop Med Hyg*. 2010;82(5):889–898. <https://doi.org/10.4269/ajtmh.2010.09-0711>
- Winkelberg S, Tasse SM, Goldstein RE et al. Evaluation of SNAP® Lepto in the diagnosis of leptospirosis infections in dogs: twenty two clinical cases. *Int J Appl Res Vet Med*. 2015;13(3):193–198
- Wynwood SJ, Graham GC, Weier SL et al. Leptospirosis from water sources. *Pathog Glob Health*. 2014;108(7):334–338. <https://doi.org/10.1179/2047773214Y.0000000156>